

Product Information

**AMPK ($\alpha 1/\beta 1/\gamma 1$), active, His-tagged, human
PRECISIO® Kinase
recombinant, expressed in *Sf9* cells**

Catalog Number **A1233**

Storage Temperature -70°C

Synonyms:

$\alpha 1$: PRKAA1, MGC33776, MGC57364

$\beta 1$: PRKAB1, AMPK, HAMPKb, MGC17785

$\gamma 1$: PRKAG1, AMPKG, MGC8666

Product Description

AMPK is a heterotrimer protein kinase consisting of an α catalytic subunit, and non-catalytic β and γ subunits. AMPK is an important energy-sensing enzyme that monitors cellular energy status.¹ In response to cellular metabolic stresses, AMPK is activated, and phosphorylates and inactivates acetyl-CoA carboxylase (ACC) and 3-hydroxy 3-methylglutaryl-CoA reductase (HMGCR), key enzymes involved in regulating fatty acid and cholesterol biosynthesis.²

This recombinant full-length human AMPK (combination of $\alpha 1/\beta 1/\gamma 1$ subunits) was expressed by baculovirus in *Sf9* insect cells using C-terminal His tags. The gene accession numbers for the three subunits ($\alpha 1/\beta 1/\gamma 1$) are NM 006251, NM 006253, and NM002733, respectively. It is supplied in 50 mM sodium phosphate, pH 7.0, 300 mM NaCl, 150 mM imidazole, 0.1 mM PMSF, 0.2 mM DTT, and 25% glycerol.

Molecular mass:

$\alpha 1$ ~68 kDa

$\beta 1$ ~38 kDa

$\gamma 1$ ~40 kDa

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at -70°C is recommended. After opening, aliquot into smaller quantities and store at -70°C . Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Typical Lot:

Purity: $\geq 70\%$ (SDS-PAGE, densitometry)

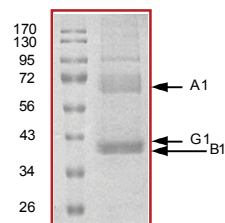
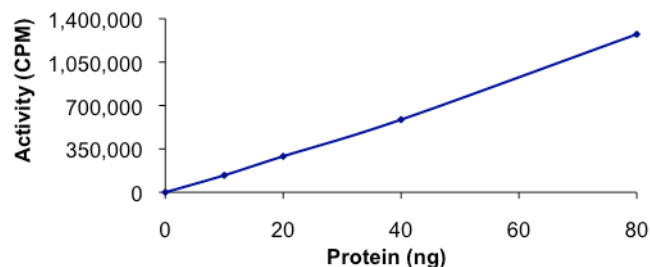


Figure 2.

Specific Activity of Typical Lot:

626–848 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl_2 , 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/ μL BSA and 5% glycerol solution.

Kinase Solution – Dilute the active AMPK ($\alpha 1/\beta 1/\gamma 1$) (0.1 $\mu\text{g}/\mu\text{l}$) with Kinase Dilution Buffer to the desired concentration.

Note: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active AMPK ($\alpha 1/\beta 1/\gamma 1$) kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μl aliquots at $-20\text{ }^{\circ}\text{C}$.

$\gamma\text{-}^{32}\text{P}$ -ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of $\gamma\text{-}^{32}\text{P}$ -ATP (1 mCi/100 μl). Store in 1 ml aliquots at $-20\text{ }^{\circ}\text{C}$.

Substrate Solution – Dissolve the synthetic peptide substrate (HMRSAMSGSLHLVKRR) in water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ^{32}P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

1. Thaw the active AMPK ($\alpha 1/\beta 1/\gamma 1$), Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The $\gamma\text{-}^{32}\text{P}$ -ATP Assay Cocktail may be thawed at room temperature.
2. In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl :
 - 10 μl of Kinase Solution
 - 5 μl of Substrate Solution
 - 5 μl of cold water ($4\text{ }^{\circ}\text{C}$)
3. Set up a blank control as outlined in step 2, substituting 5 μl of cold water ($4\text{ }^{\circ}\text{C}$) for the Substrate Solution.
4. Initiate each reaction with the addition of 5 μl of the $\gamma\text{-}^{32}\text{P}$ -ATP Assay Cocktail, bringing the final reaction volume to 25 μl . Incubate the mixture in a water bath at $30\text{ }^{\circ}\text{C}$ for 15 minutes.
5. After the 15 minute incubation, stop the reaction by spotting 20 μl of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

6. Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~ 10 minutes each.
7. Set up a radioactive control to measure the total $\gamma\text{-}^{32}\text{P}$ -ATP counts introduced into the reaction. Spot 5 μl of the $\gamma\text{-}^{32}\text{P}$ -ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

$$\text{SR} = \frac{\text{cpm of 5 } \mu\text{l of } \gamma\text{-}^{32}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

cpm – value from control (step 7)
nmole – 1.25 nmole (5 μl of 250 μM ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

$$\text{nmole/min/mg} = \frac{\Delta\text{cpm} \times (25/20)}{\text{SR} \times \text{E} \times \text{T}}$$

SR = specific radioactivity of the ATP (cpm/nmole ATP)

Δcpm = cpm of the sample – cpm of the blank (step 3)

25 = total reaction volume

20 = spot volume

T = reaction time (minutes)

E = amount of enzyme (mg)

References

1. Minokoshi, Y. et al., AMP-kinase regulates food intake by responding to hormonal and nutrient signals in the hypothalamus. *Nature*, **428**, 569-574, (2004).
2. Hardie, D.G. et al., The AMP-activated protein kinase-fuel gauge of the mammalian cell? *Eur. J. Biochem.*, **246**, 259-73 (1997).

PRECISIO is a registered trademark of Sigma-Aldrich Co. LLC.

JR,SV,MAM 04/12-1