

Product Information

Lysozyme from Chicken Egg White

Powder or Granules, $\geq 39,000$ units/mg protein**L6876**

Product Description

CAS Registry Number: 12650-88-3

Enzyme Commission (EC) Number: 3.2.1.17

Synonym: Muramidase, Lysozyme c,
Mucopolysaccharide *N*-acetylmuramoylhydrolase

Lysozyme is a single chain polypeptide of 129 amino acids cross-linked with four disulfide bridges.¹ Lysozyme hydrolyzes $\beta(1\rightarrow4)$ linkages between *N*-acetyl-muraminic acid and *N*-acetyl-D-glucosamine residues in peptidoglycan and between *N*-acetyl-D-glucosamine residues in chitodextrin.^{2,3}

Lysozyme is often used for lysing bacterial cells by hydrolyzing the peptidoglycan present in the cell walls:

- Gram-positive cells are quite susceptible to this hydrolysis, as their cell walls have a high proportion of peptidoglycan.
- Gram-negative bacteria are less susceptible due to the presence of an outer membrane and a lower proportion of peptidoglycan. However, these cells may be hydrolyzed more easily in the presence of EDTA that chelates metal ions in the outer bacterial membrane.^{4,5}

This highly purified enzyme preparation has been used in mass spectrometry as a protein mass calibration standard and in structural studies of proteins.⁶⁻⁸ Several theses⁹⁻¹¹ and dissertations¹²⁻²⁴ have cited use of product L6876 in their protocols.

Molecular mass: 14,307 Da (calculated from amino acid sequence)²⁵

Isoelectric point (pI): 11.35²⁶

Extinction coefficients:

- $E_{1\%}^{1\text{cm}}$ (281.5 nm): 26.4 in 0.1 M KCl²⁷
- $E_{\text{mM}}^{1\text{cm}}$ (280 nm): 36²⁸

Optimal pH

- The activity of lysozyme is a function of both pH and ionic strength.²⁸
- The enzyme is active over a broad pH range (6.0-9.0).²⁸
- At pH 6.2, maximal activity is observed over a wider range of ionic strengths (0.02 M - 0.100 M) than at pH 9.2 (0.01 M - 0.06 M).²⁸

Inhibitors

- Indole derivatives (which bind to and distort the active site)²⁹
- Imidazole (which induces the formation of a charge-transfer complex)²⁹
- Surface-active agents such as sodium dodecyl sulfate (SDS), sodium dodecanate, and dodecyl alcohol
 - Other compounds of these types with carbon chains of 12 or more carbons in length will also inhibit lysozyme.³⁰

Substrates

The natural substrate for lysozyme is the peptidoglycan layer of bacterial cell walls. However, various low molecular mass substrates, including murein degradation products, as well as synthetic compounds, have been used for different photometric, isotopic, and immunological lysozyme assays.³¹

The following low molecular mass lysozyme substrates are available:

- 4-Methylumbelliferyl β -D-*N,N,N'*-triacetylchitotriose (Cat. No. M5639, a fluorogenic substrate)
- 4-Nitrophenyl β -D-*N,N,N'*-triacetylchitotriose (Cat. No. N8638, a chromogenic substrate)

Product

This lysozyme preparation is purified from chicken egg white.

Lysozyme activity: $\geq 39,000$ units/mg protein

Unit definition: One unit will produce a change in A_{450} of 0.001 per minute at pH 6.24 at 25 °C, using a suspension of *Micrococcus lysodeikticus* as substrate, in a 2.6 mL reaction mixture (1 cm light path).

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Preparation Instructions

For *E. coli* cell lysis, use a freshly prepared lysozyme solution (10 mg/mL) in 10 mM Tris-HCl, pH 8.0.³²

The product is also soluble in water (10 mg/mL) yielding a clear to slightly hazy colorless solution. Aqueous solutions should retain activity for at least one month when stored between 2-8 °C.

Storage/Stability

The product, as supplied, should be stored at room temperature.

Procedure

The following procedure is for the lysis of *E. coli*. It may be used as a guideline for other species. The optimal pH for *E. coli* cell lysis is 8.0 ± 0.1 .¹⁶

1. Incubate *E. coli* (strain ATCC 37017) bearing the pBR322 plasmid overnight in Terrific Broth (Cat. No. T0918) with 25 µg/mL tetracycline (Cat. No. T3383) and 25 µg/mL ampicillin (Cat. No. A9518).
2. Centrifuge 1-2 mL samples of the overnight culture.
3. Resuspend the pellets in 350 µL of STET buffer (10 mM Tris-HCl, pH 8.0, with 0.1 M NaCl, 1 mM EDTA, and 5% [w/v] TRITON® X-100).
4. Add 25 µL of a freshly prepared lysozyme solution (10 mg/mL in 10 mM Tris-HCl, pH 8.0).
5. Mix by vortexing for 3 seconds.
6. Incubate the lysis mixture for 30 minutes at 37 °C.
7. After incubation, place the tube containing the lysis mixture in a boiling water bath for exactly 40 seconds.

8. Centrifuge the lysis mixture at $14,000 \times g$.
9. Remove the pellet (cell debris) from the tube using a sterile toothpick.
10. Plasmid DNA from the supernatant may then be purified and analyzed.

References

1. Jollès, P., Angew. Chem. Int. Ed. Engl., 8(4), 227-239 (1969).
2. Rupley, J.A., Biochim. Biophys. Acta, 83(3), 245-255 (1964).
3. Holler, E., et al., Biochemistry, 14(11), 2377-2385 (1975).
4. Schütte, H., and Kula, M.R., Biotechnol. Appl. Biochem., 12(6), 599-620 (1990).
5. Vázquez-Laslop, N. et al., J. Bacteriol., 183(8), 2399-2404 (2001).
6. Galvani, M. et al., Electrophoresis, 22(10), 2058-2065 (2001).
7. Abgar, S. et al., Eur. J. Biochem., 267(19), 5916-5925 (2000).
8. Sethuraman, A. et al., Proteins, 56(4), 669-678 (2004).
9. Chen, Timothy Linghau, "Investigating the in vivo Folding Kinetics and Thermodynamics of Phosphoglycerate Kinase". University of Illinois Urbana-Champaign, B.S. thesis, p. 27 (2016).
10. Jin, Minfeng, "Sustained release of lysozyme encapsulated in zein micro- and nanocapsules". University of Tennessee Knoxville, M.S. thesis, p. 37 (2008).
11. Blackwell, Susan, "Characterization of the Saccharomyces cerevisiae RAD5 Gene and Protein". University of Saskatchewan, M.Sc. thesis, p. 27 (2013).
12. Hinshelwood, Justin, "Structural Studies of factor B of the Alternative Pathway of the Complement System". University College London, Ph.D. dissertation, p. 97 (2000).
13. Gokarn, Yatin R., "Hydrodynamic behavior and thermal stability of a PEGylated protein: Studies with hen egg lysozyme". University of New Hampshire, Ph.D. dissertation, p. 21 (2003).
14. Bartling, Karsten, "Apoferritin Crystallization in Relation to Eye Cataract". Georgia Institute of Technology, Ph.D. dissertation, p. 85 (2006).
15. Leigh, Brian, "Electron transfer through organic and biological molecules". California Institute of Technology, Ph.D. dissertation, p. 70 (2009).
16. Shah, Umang Vinubhai, "3D Nanotemplates for Protein Crystallisation". Imperial College London, Ph.D. dissertation, p. 104 (2012).

17. Vance, Steven J., "The relationship between structure and function in natural surfactant proteins". University of Glasgow, Ph.D. dissertation, p. 90 (2012).
18. Ward, Catherine Louise, "The cellular consequences of FUS/TLS depletion: a loss of function model for amyotrophic lateral sclerosis". University of Massachusetts Graduate School of Biomedical Sciences, Worcester, Ph.D. dissertation, p. 177 (2014).
19. Thyparambil, Aby Abraham, "Development, validation, and application of analytical methods for characterizing adsorbed protein orientation, conformation, and bioactivity". Clemson University, Ph.D. dissertation, pp. 118, 163, 236, 258 (2015).
20. Delacher, Michael, "Transcriptional Control of Regulatory T cells". Ruperto-Carola University Heidelberg, Dr. rer. nat. dissertation, p. 176 (2016).
21. Busse, Frederik, "Mechanisms of Picosecond Infrared Laser Desorption Ionization". University of Hamburg, Ph.D. dissertation, p. 81 (2019).
22. Mason, Bethany Jane, "Functional analysis of the F-box protein Fbx17". University of Cambridge, Ph.D. dissertation, p. 63 (2019).
23. Moreau, David Wayne, "Ice Formation and Solvent Nanoconfinement in Protein Crystallography". Cornell University, Ph.D. dissertation, pp. 15, 41, 115 (2020).
24. Ramos, João Carlos Moreno, "Protein and water dynamics at the atomic level". University of Copenhagen, Ph.D. dissertation, pp. 72, 114 (2022).
25. Canfield, R.E., J. Biol. Chem., 238(8), 2698-2707 (1963).
26. Wetter, L.R., and Deutsch, H.F., J. Biol. Chem., 192(1), 237-242 (1951).
27. Aune, K.C., and Tanford, C., Biochemistry, 8(11), 4579-4585 (1969).
28. Davies, R.C., et al., Biochim. Biophys. Acta, 178(2), 294-305 (1969).
29. Swan, I., J. Mol. Biol., 65(1), 59-62 (1972).
30. Smith, G., and Stoker, C., Arch. Biochem., 21(2), 383-394 (1949).
31. Höltje, J.V., EXS, 75, 105-110 (1996).
32. Sambrook, J. et al. (eds.), Molecular Cloning, A Laboratory Manual. Cold Spring Harbor Laboratory Press, (Cold Spring Harbor, NY), p 1.29 (1989).

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23193256 Rev 03/25

